# **Reactivos GPL**

Barcelona, España

# Store at: +2+8°C.

-UREA-37-

# UREA

o-Phthalaldehyde 37°C. Colorimetric.

Presentation:

CE

Cod. SU040 CONT: R1 1 x 125 mL.+ R2 1 x 125 mL.+ Cal. 1 x 5 mL.

# Quantitative determination of urea.

Only for in vitro use in clinical laboratory (IVD)

# TEST SUMMARY

Urea in the sample reacts with o-fhthalaldehyde in acid medium forming a coloured complex that can be measured by spectrophotometry:

Urea + o-fhthalaldehyde  $\xrightarrow{H^+}$  Isoindoline

The intensity of the color formed is proportional to the urea concentration in the sample<sup>1</sup>

# **REAGENTS COMPOSITION**

R.1	o-Phthalaldehyde	4.8 mmol/L
R.2	Borate solution	87 mmol/L
Calibrator	Urea aqueous primary standard	50 mg/dL

# **REAGENT PREPARATION AND STABILITY**

All the reagents are ready to use.

All the components of the kit are stable until the expiration date on the label when stored tightly closed at 2-8°C, protected from light and contamination prevented during their use.

Do not use reagents over the expiration date.

Urea Cal.: Once open is stable up to 1 month when stored tightly closed at 2-8°C, protected from light and contaminations prevented during their use. Signs of reagent deterioration:

## Presence of particles and turbidity

Blank absorbance (A) at 510 nm > 0.20

All the reagents of the kit are stable up to the end of the indicated month and year of expiry. Stored at tightly closed at 2-8°C,. Do not use reagents over the expiration date.

### **SPECIMEN**

Serum or heparinized plasma<sup>1</sup>: Do not use ammonium salts or fluoride as anticoagulants.

Urine<sup>1</sup>: Dilute sample 1/50 in distilled water. Mix. Multiply results by 50 (dilution factor). Preserve urine samples at pH < 4. Urea is stable at 2-8°C for 5 days.

# MATERIAL REQUIRED BUT NOT PROVIDED

- Spectrophotometer or colorimeter measuring at 510 nm.
- Matched cuvettes 1.0 cm. light path.
- General laboratory equipment (Note 1).

### TEST PROCEDURE A) KINETIC METHOD

Pipette into a cuvette: 1

	Blank	Standard	Sample			
R 1 (mL)	1.0	1.0	1.0			
Standard <sup>(Note 2-3)</sup> (µL)		50				
Sample (µL)			50			
2. Mix, wait 1 minute and add:						
R 2 (mL)	1.0	1.0	1.0			

Mix, incubate at 37°C and read the absorbance after 1 minute (A1) and after 2 minutes (A2).

Calculate the increase of the absorbance  $\Delta A = A_2 - A_1$ .

# CALCULATIONS

Urea mg/dL =  $\frac{(\Delta A) \text{Sample}}{(\Delta A)^2} \times 50$  (Calibrator conc.)

# (AA)Calibrator

# **B) END POINT**

1. Pipette into a cuvette:					
	Blank	Standard	Sample		
R 1 (mL)	1.0	1.0	1.0		
Standard <sup>(Note 2-3)</sup> (µL)		25			
Sample (µL)			25		
2. Mix, wait 1 min and add:					
R 2 (mL)	1.0	1.0	1.0		

3 Mix and incubate 15 min at 37°C

4 Read the absorbance (A) against the Blank.

(A)Calibrator

Control sera are recommended to monitor the performance of the

If control values are found outside the defined range, check the instrument.

actions if controls do not meet the acceptable tolerances.

# **REFERENCE VALUES,1**

Serum or Plasma				
15-45 mg/dL	2.5-7.5 mmol/L			
Urine				
20-35 g/24 h				

### (These values are for orientation purpose)

It is suggested that each laboratory establish its own reference range. CLINICAL SIGNIFICANCE

Urea is the final result of the metabolism of proteins; it is formed in the liver from its destruction.

Elevated urea can appear in blood (uremia) in: diets with excess of proteins, renal diseases, heart failure, gastrointestinal hemorrhage, dehydration or renal obstruction<sup>1,6,7</sup>.

Clinical diagnosis should not be made on a single test result; it should integrate clinical and other laboratory data

# REAGENT PERFORMANCE

<u>Measuring Range</u>: From detection limit of 0.70 mg/dL. to linearity limit of 200 mg/dL., under the described assay conditions. If results obtained were greater than linearity limit, dilute the sample  $\frac{1}{2}$ with NaCl 9 g/L. and multiply result by 2.

Precision

	Intra-assay (n=20)		Inter-assay (n=20)	
Mean (mg/dL)	43.2	145	41.9	147
SD	1.51	1.10	0.80	2.83
CV (%)	3.49	0.76	1.92	1.91

Sensitivity: 1 mg/dL. = 0.00459 A

## Accuracy:

Results obtained GPL reagents did not show systematic differences when compared with other commercial reagents.

The results of the performance characteristics depend on the analyzer used.

# INTERFERING SUBSTANCES

# Interference:

It is recommended to use heparin as anticoagulant. Do not use ammonium salts or fluoride1

A list of drugs and other interfering substances with urea determination has been reported by Young et.  $al^{2.3}$ .

### NOTES

Glassware and distilled water must be free of ammonia and 1. ammonium salts<sup>1</sup>

- 2. Calibration with the aqueous standard may cause a systematic error in automatic procedures. In these cases, it is recommended to use a serum Calibrator.
- 3. Use clean disposable pipette tips for its dispensation.

# BIBLIOGRAPHY

- Kaplan A. Urea. Kaplan A et al. Clin Chem The C.V. Mosby Co. St Louis. Toronto. Princeton 1984; 1257-1260 and 437 and 418. Young DS. Effects of drugs on Clinical Lab. Tests, 4th ed AACC Press, 1995. 1.
- 2.
- 3
- Young DS. Effects of disease on Clinical Lab. Tests, 4th ed AACC 2001. Burtis A et al. Tietz Textbook of Clinical Chemistry, 3rd ed AACC 1999. Tietz N W et al. Clinical Guide to Laboratory Tests, 3rd ed AACC 1995. 4. 5



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Procedure

CALCULATIONS

(A)Sample - x 50 (Calibrator conc.) Urea mg/dL =

10 mg/L urea BUN divided by 0.466 = 21 mg/L urea = 0.36 mmol/L urea<sup>1</sup>.

Conversion factor: mg/dL x 0.1665 = mmol/L.

# **OUALITY CONTROL**

procedure, Normal and Pathological.

reagents and calibrator for problems.

Serum controls are recommended for internal quality control. Each laboratory should establish its own Quality Control scheme and corrective